



Complete Genome Sequences of *Thermus* Strains Isolated from Senami Hot Spring in Japan

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ABSTRACT Extremely thermophilic strains belonging to the genus *Thermus* were isolated from Senami Hot Spring in Japan. Here, I report the complete genome sequences of five *Thermus thermophilus* strains and one *Thermus brockianus* strain, which were obtained by combining Oxford Nanopore long-read and DNBSEQ or Illumina short-read sequencing data.

*T*hermus aquaticus is the type species of the genus *Thermus*, which was first isolated from Yellowstone National Park in the United States in 1969 (1). At almost the same time, *Thermus thermophilus* was isolated from Mine Hot Spring in Japan (2, 3). Due to its high-temperature adaptation at around 70°C, *Thermus* has great biotechnological potential as a source of thermophilic enzymes, as exemplified by *Taq* DNA polymerase (4). Since the discovery of *T. aquaticus*, many *Thermus* strains have been isolated from thermal areas worldwide (5–11).

I collected boiling water samples at Senami Hot Spring (38.2139 N, 139.4438 E) in Japan. Samples were spread over *Thermus* medium (0.4% [wt/vol] yeast extract, 0.8% [wt/vol] peptone, 0.2% [wt/vol] NaCl) agar plates (1.6% [wt/vol]) containing 0.4 mM MgCl₂ and 0.35 mM CaCl₂ (*Thermus* MC medium) (10). After incubation at 75°C overnight, dozens of well-separated single colonies were isolated, and colony PCR was conducted to analyze the 16S rRNA gene using the *Thermus*_1F and *Thermus*_1521R primer sets (12). I selected five *T. thermophilus* strains (≥99.8% 16S rRNA identities to HB8^T [[TTH_RS00710](#)]) and one *T. brockianus* strain (99.9% 16S rRNA identity to YS38^T [[NR_036983.1](#)]) for whole-genome analysis.

To prepare genomic DNA, cells were aerobically grown in 5 mL of *Thermus* MC medium at 75°C for 24 h. Genomic DNA was purified using a blood and cell culture DNA midikit (Qiagen; catalog number 13323). For long-read sequencing, unsheared genomic DNA (1 µg) was treated with a short-read eliminator kit (Circulomics; catalog number SS-100-121-01) to remove fragments of <10 kbp, and a library was constructed using a ligation sequencing kit (Oxford Nanopore Technologies [ONT]; catalog number SQK-LSK109). Sequencing was performed with a GridION X5 system on a FLO-MIN106 R9.41 revD flow cell (ONT). Base calling was conducted using Guppy v.4.0.11. The raw data (Table 1) were filtered (Q > 10; length, >1,000 bases) using NanoFilt v.2.7.1 (13). For short-read sequencing of SNM1-1, SNM3-3, and SNM1-7, a library was constructed using an MGIEasy FS PCR Free DNA library preparation set (MGI Tech Co., Ltd; catalog number 1000013455) with a ~400- to 500-bp insert. Paired-end sequencing (2 × 150 bases) was then performed on a DNBSEQ-400 instrument (MGI). For SNM6-6, SNM7-6, and SNM4-1, the DNA prep kit (Illumina; catalog number 20018704) was used to generate paired-end libraries with approximately 350-bp inserts. Sequencing was performed using a MiSeq reagent kit v.2 (300 cycles) with 256-bp reads. These raw data were filtered (Q > 30; length, >20 bases) using fastp v.0.20.1 (14) (Table 1). Default parameters were used for all software. The trimmed long- and short-read data were assembled using Unicycler v.0.4.8 (15), and the assembly was polished using Pilon v.1.24 (16).

All strains contained a single circular chromosome and multiple circular plasmids. Circularity was confirmed via Unicycler. Automatic annotation was conducted using DFAST v.1.2.15 (17); genomic features are summarized in Table 1.

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TABLE 1 Strains sequenced in this study

Strain	BioSample accession no.	Chromosome or plasmid	Short read			Long read			SRA accession no.	Total length (Mb)	GC content (%)	avg read depth (X)	No. of coding sequences	GenBank accession no.
			No. of paired end reads	Total length (Mb)	SRA accession no.	No. of reads	N_{50} (bases)							
<i>T. thermophilus</i> SNM1-1	SAMD00442895	Chromosome pTthSNM1-1b pTthSNM1-1c pTthSNM1-1d pTthSNM1-1e pTthSNM1-1f pTthSNM1-1g	12,080,398	1.812	DRR346611	676,877	6,660	2,160	DRR346617	2,126,669	69.31	107	2,371	AP025596 AP025597 AP025598 AP025599 AP025600 AP025601 AP025602
<i>T. thermophilus</i> SNM3-3	SAMD00442896	Chromosome pTthSNM3-3b pTthSNM3-3c pTthSNM3-3d	5,407,329	811	DRR346612	1,066,704	3,727	2,995	DRR346618	1,971,781	69.34	676	2,113	AP025609 AP025610 AP025611 AP025612
<i>T. thermophilus</i> SNM6-6	SAMD00442897	Chromosome pTthSNM6-6b pTthSNM6-6c pTthSNM6-6d	963,253	226	DRR346613	1,459,467	3,609	2,897	DRR346619	1,945,718	69.41	136	2,087	AP025613 AP025614 AP025615 AP025616
<i>T. thermophilus</i> SNM7-6	SAMD00442898	Chromosome pTthSNM7-6b pTthSNM7-6c pTthSNM7-6d	989,094	227	DRR346614	1,608,666	1,807	2,619	DRR346620	1,848,270	69.53	123	1,978	AP025617 AP025618 AP025619 AP025620
<i>T. thermophilus</i> SNM1-7	SAMD00442899	Chromosome pTthSNM1-7b pTthSNM1-7c pTthSNM1-7d pTthSNM1-7e pTthSNM1-7f	4,871,052	731	DRR346615	1,096,885	13,982	3,978	DRR346621	1,870,712	69.44	381	2,022	AP025603 AP025604 AP025605 AP025606 AP025607 AP025608
<i>T. brockianus</i> SNM4-1	SAMD00442900	Chromosome pTbSNM4-1b pTbSNM4-1c	1,139,199	249	DRR346616	1,088,755	4,184	3,050	DRR346622	2,046,081	66.96	214	2,181	AP025593 AP025594 AP025595

Data availability. All six *Thermus* strains are associated with BioProject PRJDB12526.

The BioSample accession numbers and accession numbers for genome sequences and raw sequencing data are available in Table 1.

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REFERENCES

1. Brock TD, Freeze H. 1969. *Thermus aquaticus* gen. n. and sp. n., a nonsporulating extreme thermophile. J Bacteriol 98:289–297. <https://doi.org/10.1128/jb.98.1.289-297.1969>.
2. Oshima T, Imahori K. 1971. Isolation of an extreme thermophile and thermostability of its transfer ribonucleic acid and ribosomes. J Gen Appl Microbiol 17:513–517. <https://doi.org/10.2323/jgam.17.513>.
3. Oshima T, Imahori K. 1974. Description of *Thermus thermophilus* (Yoshida and Oshima) comb. nov., a nonsporulating thermophilic bacterium from a Japanese thermal spa. Int J Syst Evol Microbiol 24:102–112. <https://doi.org/10.1099/00207713-24-1-102>.
4. Innis MA, Myambo KB, Gelfand DH, Brow MA. 1988. DNA sequencing with *Thermus aquaticus* DNA polymerase and direct sequencing of polymerase chain reaction-amplified DNA. Proc Natl Acad Sci U S A 85:9436–9440. <https://doi.org/10.1073/pnas.85.24.9436>.
5. Manaia CM, Hoste B, Gutierrez MC, Gillis M, Ventosa A, Kersters K, Da Costa MS. 1995. Halotolerant *Thermus* strains from marine and terrestrial hot springs belong to *Thermus thermophilus* (ex Oshima and Imahori, 1974) nom. rev. emend. Syst Appl Microbiol 17:526–532. [https://doi.org/10.1016/S0723-2020\(11\)80072-X](https://doi.org/10.1016/S0723-2020(11)80072-X).
6. Santos MA, Williams RA, Da Costa MS. 1989. Numerical taxonomy of *Thermus* isolates from hot springs in Portugal. Syst Appl Microbiol 12:310–315. [https://doi.org/10.1016/S0723-2020\(89\)80079-7](https://doi.org/10.1016/S0723-2020(89)80079-7).
7. Fujino Y, Kawatsu R, Inagaki F, Umeda A, Yokoyama T, Okaue Y, Iwai S, Ogata S, Ohshima T, Doi K. 2008. *Thermus thermophilus* TMY isolated from silica scale taken from a geothermal power plant. J Appl Microbiol 104:70–78. <https://doi.org/10.1111/j.1365-2672.2007.03528.x>.
8. Miyazaki K, Tomariguchi N. 2019. Complete genome sequences of *Thermus thermophilus* strains AA2-20 and AA2-29, isolated from Arima Onsen in Japan. Microbiol Resour Announc 8:e00820-19. <https://doi.org/10.1128/MRA.00820-19>.
9. Miyazaki K, Tomariguchi N, Ueno Y. 2021. Complete genome sequences of four halophilic *Thermus thermophilus* strains isolated from Arima Hot Spring in Japan. Microbiol Resour Announc 10:e00874-21. <https://doi.org/10.1128/MRA.00874-21>.
10. Miyazaki K, Moriya T, Nemoto N, Oshima T, Yura K, Bessho Y. 2021. Complete genome sequence of *Thermus thermophilus* strain HB5018, isolated from Mine Hot Spring in Japan. Microbiol Resour Announc 10:e00039-21. <https://doi.org/10.1128/MRA.00039-21>.
11. Miyazaki K, Moriya T, Tokito N, Oshima T, Yura K, Bessho Y. 2021. Complete genome sequences of *Thermus thermophilus* strains HB5002 and HB5008, isolated from Mine Hot Spring in Japan. Microbiol Resour Announc 10:e00272-21. <https://doi.org/10.1128/MRA.00272-21>.
12. Miyazaki K, Tomariguchi N. 2019. Occurrence of randomly recombined functional 16S rRNA genes in *Thermus thermophilus* suggests genetic interoperability and promiscuity of bacterial 16S rRNAs. Sci Rep 9:11233. <https://doi.org/10.1038/s41598-019-47807-z>.
13. De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.
14. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>.
15. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
16. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
17. Tanizawa Y, Fujisawa T, Kamimura E, Nakamura Y, Arita M. 2016. DFAST and DAGA: web-based integrated genome annotation tools and resources. Biosci Microbiota Food Health 35:173–184. <https://doi.org/10.12938/bmfh.16-003>.